

Application of the ¹⁵N-Gas Flux method for measuring in situ N₂ and N₂O fluxes due to denitrification in soils and comparison with the acetylene inhibition method

INTRODUCTION

The ¹⁵N tracer approaches can provide *in situ* measurements of both N₂ and N₂O, but their use has been limited to fertilised arable soils due to the need for large ¹⁵N additions in order to detect ¹⁵N₂ production against the high atmospheric N₂. An 'in house' laboratory designed and manufactured N₂ preparation instrument interfaced to a continuous flow isotope ratio mass spectrometer (CF-IRMS) can allow the analysis of ¹⁵N-N₂ with small injection volumes, improved precision and lower limit of detection. Such an instrumental advance could improve our ability for measuring denitrification in natural and semi-natural land use types. Therefore, we designed a study to:

- 1) Determine the precision and suitability of our preparative-IRMS instrumentation for measuring ¹⁵N-N₂ and ¹⁵N-N₂O at low/trace enrichment levels
- 2) Adapt the ¹⁵N Gas-Flux method for application across natural and semi-natural terrestrial ecosystems
- 3) Directly compare the validity and applicability of the ¹⁵N Gas-Flux method with the acetylene inhibition technique (AIT) for measuring *in situ* denitrification rates.

METHODS

For N₂ gas isotopic analysis an Isoprime IRMS coupled to an 'in house' built N₂ preparative interface was used (Fig 1). Headspace gas (4 µL) was injected and ratios for the m/z 28, m/z 29 and m/z 30 were recorded. For N₂O, headspace gas (*ca*. 4mL) was injected into a TraceGasTM Preconcentrator coupled to an IRMS and ratios for m/z 44, m/z 45 and m/z 46 were measured.

In situ denitrification rates in organic (OS), woodland (WL) and grassland (GL) soils were measured using static chambers (Fig 2A) according to the ¹⁵N Gas-Flux method¹. Labelled K¹⁵NO₃⁻ (98 at. %) was applied in each site (n= 5) via multiple injections into enclosed soils. Gas samples were collected at T = 1h, T = 2h and T \approx 20h for N₂ and N₂O analysis. Minimum detectable concentration (MDC) change for R29 and R30 was defined² using standards to determine if each time step sample was significantly different from ambient (T= 0 hr) and if not they were excluded from the flux calculations. The flux of N₂ and N₂O were determined ^{3, 4}.

At the same time intact soils cores were collected and incubated *in situ* with and without the addition of C_2H_2 according to the AIT approach for the determination of denitrification rates (Fig 2 B)⁵.



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Figure 2: ¹⁵N Gas flux chamber (A) and intact soil scores with C_2H_2 amendment (B)



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Instrument stability checks showed standard deviation fits better than 0.05 ‰ for both gases. Precision of the instrument was better than 0.08 ‰ and 0.3 ‰ for δ^{15} N-N₂ and δ^{15} N-N₂O gases, respectively.

The minimum detectable flux rates were 4 μ g N m⁻² h⁻¹ and 0.2 ng N m⁻² h⁻¹ for N₂ and N₂O, respectively, which is a significant improvement compared to earlier studies. The improved precision for both allowed us to quantify denitrification with low ¹⁵N enrichment under *in situ* conditions, which was not possible earlier.

The evolved N₂ and N₂O in the chamber headspace increased linearly from 1 to 20 hours (Fig 3). We calculated flux rates by applying linear regression (when $r^2 > 0.95$) between 1 and 20 hours using only those time points that were above the MDC values.

The total denitrification rate measured using the ¹⁵N Gas flux (range: 2.4 - 416.6 µg N m⁻² h⁻¹) and the C₂H₂ methods (range: 0.5 - 325.2 μ g N m⁻² h⁻¹) followed a similar trend across the sites (Pearson; r = 0.581, n = 25, p < 0.01) (Fig 4). However, denitrification rates measured using the ¹⁵N Gas flux method were between 3 and 5 times higher than the denitrification rates with the AIT method.

Bulk N₂O emission rates measured using the headspace samples from the chambers and no-C₂H₂ amended cores exhibited a similar trend across sites; however, the N₂O/N₂+N₂O ratios differed between the two methods (Fig 4). The N₂O/N₂ + N₂O ratio measured using the ¹⁵N Gas flux method was low (range: 0.03 to 13%) compared to the AIT (range: 50 to 60%). The reason for this discrepancy is that the AIT cannot discriminate N₂O sources to constrain the ratio to denitrification only and an incomplete inhibition of N₂O reduction due to diffusion constraints of C_2H_2 in soil cores.

The improved precision for both ¹⁵N-N₂ and ¹⁵N-N₂O analyses allows the quantification of in situ denitrification rates with low ¹⁵N enrichment in natural and semi-natural ecosystems.

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RESULTS AND DISCUSSION

CONCLUSION

REFERENCES



using the ¹⁵N Gas flux method.